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## DOCUMENTATION PAGE

Form Approved  
OMB No. 0704-0188

1a. REPORT SECURITY CLASSIFICATION (U)		1b. RESTRICTIVE MARKINGS NA	
2a. SECURITY CLASSIFICATION AUTHORITY NA		3. DISTRIBUTION/AVAILABILITY OF REPORT Distribution unlimited	
2b. DECLASSIFICATION/DOWNGRADING SCHEDULE NA			
4. PERFORMING ORGANIZATION REPORT NUMBER(S) Princeton University		5. MONITORING ORGANIZATION REPORT NUMBER(S) NA	
6a. NAME OF PERFORMING ORGANIZATION Princeton University	6b. OFFICE SYMBOL (If applicable) NA	7a. NAME OF MONITORING ORGANIZATION Office of Naval Research	
6c. ADDRESS (City, State, and ZIP Code) Princeton, NJ 08544		7b. ADDRESS (City, State, and ZIP Code) 800 N. Quincy St. Arlington, VA 22217-5000	
8a. NAME OF FUNDING/SPONSORING ORGANIZATION Office of Naval Research	8b. OFFICE SYMBOL (If applicable) ONR	9. PROCUREMENT INSTRUMENT IDENTIFICATION NUMBER N00014-86-K-0396-P00001	
8c. ADDRESS (City, State, and ZIP Code) 800 N. Quincy St. Arlington, VA 22217-5000		10. SOURCE OF FUNDING NUMBERS PROGRAM ELEMENT NO. 61153N	PROJECT NO. RR04108
		TASK NO. 441k818	WORK UNIT ACCESSION NO.
11. TITLE (Include Security Classification) (U) Lipid Dependent Mechanisms of Protein Pump Activity			
12. PERSONAL AUTHOR(S) Gruner, Sol M.			
13a. TYPE OF REPORT Annual	13b. TIME COVERED FROM 8/87 TO 7/88	14. DATE OF REPORT (Year, Month, Day) 88/7/11	15. PAGE COUNT
16. SUPPLEMENTARY NOTATION			
17. COSATI CODES FIELD GROUP SUB-GROUP 08		18. SUBJECT TERMS (Continue on reverse if necessary and identify by block number) Lipid effect; protein pumps; membrane protein	
19. ABSTRACT (Continue on reverse if necessary and identify by block number) It has been proposed that membrane protein pump function is modulated by the lipid monolayer spontaneous curvature, an elastic parameter of the monolayers related to the tendency for the lipids to form non-bilayer phases. [Gruner, Proc. Natl. Acad. Sci., USA 82 (1985) 3665]. To test this hypothesis $Ca^{2+}$ ATPase from rabbit sarcoplasmic reticulum has been reconstituted into vesicles of different lipids with well characterized values of the spontaneous curvature. The activity is found to correlate with the spontaneous curvature and not with the specific lipids used to achieve a given value of spontaneous curvature. Progress is also reported on the development of alternative methods of measuring spontaneous membrane curvature. <i>radius of curvature</i>			
20. DISTRIBUTION/AVAILABILITY OF ABSTRACT <input checked="" type="checkbox"/> UNCLASSIFIED/UNLIMITED <input type="checkbox"/> SAME AS RPT <input type="checkbox"/> DTIC USERS		21. ABSTRACT SECURITY CLASSIFICATION (U)	
22a. NAME OF RESPONSIBLE INDIVIDUAL Dr. Igor Vodyanoy		22b. TELEPHONE (Include Area Code) 202-696-4056	22c. OFFICE SYMBOL ONR

DD Form 1473, JUN 86

Previous editions are obsolete.

SECURITY CLASSIFICATION OF THIS PAGE

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Date: 28 June 1988

## Progress Report on Contract N00014-86-K-0396-P00001

Principal Investigator: Dr. Sol M. Gruner

Contractor: Princeton University

Contract Title: Lipid Dependent Mechanisms of Protein Pump Activity

**INTRODUCTION:**

The physical parameter of lipid bilayers which modulate the activity of integral protein membrane are not well understood. It has been proposed that the value of an elastic parameter of the lipid monolayers, known as the monolayer spontaneous radius of curvature,  $R_0$ , is important to the activity of certain membrane proteins [1]. The overall goal of this research is to test this hypothesis. This involves three objectives:

- 1) Develop methods of measuring the spontaneous radius of curvature of lipid membranes and better understand the physical effects of varying  $R_0$ .
- 2) Correlate the activity of a well characterized pump protein with the  $R_0$  value of the imbedding bilayer. If  $R_0$  is important, then the pump activity should correlate with  $R_0$ , not with the specific lipid mix used to achieve a given  $R_0$  value.
- 3) Determine if bacterial membranes act to keep  $R_0$  fixed when the bacteris are forced to alter their membrane lipid composition.

**PROGRESS:**

Objective 1). A method of measuring  $R_0$  for pure lipids (e.g., in the absence of protein has been developed [2,3]. However, it is desirable to develop methods which operate in bilayers containing protein and over a wider range of  $R_0$  values. There is evidence that the  $^2\text{H}$ -NMR quadropole splitting,  $\Delta Q$  of deuterated lipid chains may correlate with  $R_0$  [4]. Toward this end, the use of perdueterated compounds as  $R_0$  probes was investigated. Relatively inexpensive lipid analogs, such as perdeuterated alkanols were found to strongly perturb membranes at levels required for good NMR signals. In consequence, perdeuterated phospholipid was recently synthesized. We are just now beginning to correlate the  $R_0$

values measured by the x-ray technique to  $\Delta Q$ . This work is being done in collaboration with Drs. P. Cullis and C. Tilcock of the University of British Columbia.

Work has also been performed on understanding the physics of  $R_0$  variation in membranes. An apparatus has been constructed which allows x-ray diffraction studies of lipid suspensions at pressures of up to 1000 bar. As expected,  $R_0$  appears to be very pressure sensitive. A note describing the pressure effects will be submitted for publication in the near future.

Objective 2). In collaboration with Drs. A. Janoff and M. Jaworsky of the Liposome Co.,  $\text{Ca}^{++}$  ATPase from rabbit sarcoplasmic reticulum has been reconstituted in vesicles of binary mixtures of the lipids DOPE, DOPE-Me and DOPC. For each mixture, the pump activity (moles  $\text{Ca}^{+2}$  pumped/moles ATP hydrolyzed) and the  $R_0$  value of the binary lipid mixture were measured. It was found that the activity correlated simple with the measured  $R_0$  values, that is to say, when pump activity was graphed against  $R_0$ , all points fell on a simple smooth curve. Different binary mixtures which had the same  $R_0$  value yielded similar pump activities. Control experiments are being done to see if the correlation is due to systematic variation in the vesicle sizes with composition. A publication describing these results will soon be prepared.

Objective 3). Lipid extracts of bacteria grown under conditions which cause the lipid composition to vary were examined to see if  $R_0$  was being held constant. Unfortunately, we have been only marginally successful in measuring  $R_0$  values on these lipids because the  $R_0$  values appear to be outside the range of our present techniques (whence, objective #1). In consequence, we are switching to another strain of bacteria which, we expect, will yield small  $R_0$  values. These cultures are being grown now. These experiments are done in collaboration with Dr. R. McElhaney of the University of Alberta and Dr. G. Lindblom of the University of Umea, Sweden.

#### REFERENCES CITED:

- [1] S.M. Gruner, Proc. Natl. Acad. Sci. *82* (1985) 3665.
- [2] S.M. Gruner, V.A. Parsegian and R.P. Rand, Faraday Discuss. Chem. Soc. *81* (1986) 29.
- [3] S.M. Gruner, M.W. Tate, G.L. Kirk, P.T.C. So, D.C. Turner, D.T. Keane, C.P.S. Tilcock and P.R. Cullis, Biochem. *27* (1988) 2853.



Availability Codes	
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- [4] P.R. Cullis, M.J. Hope and C.P.S. Tilcock, *Chem. Phys. Lipids* 40 (1986) 127.

#### WORK PLAN:

- 1) Sufficient quantities of perdeuterated lipid are in hand. We intend to proceed immediately to correlate the  $^2\text{H}$ -NMR of these materials with  $R_0$  values derived by x-ray diffraction.
- 2) The  $\text{Ca}^{++}$  ATPase will next be reconstituted in a broader range of lipids. Systematic control studies on the vesicle sizes will also be done to insure that the observed correlation is not simply a vesicle size effect. It is also known that  $R_0$  is a sensitive function of pressure. We will map out how  $R_0$  changes with pressure for representative lipid mixtures. It would be of interest to see if the correlation of pump activity varies with pressure as expected from the variation in  $R_0$  due to composition.
- 3) We hope to have enough lipid extracts from new bacterial cultures by late summer to proceed with  $R_0$  measurements. We will first use the x-ray diffraction measurement procedure. If difficulties are encountered with this, we will have to wait until the NMR method is refined.
- 4) Investigation of the effects of lipid spontaneous curvature will be extended to oligopeptide membrane channels, as discussed in the continuation application for this contract.

INVENTIONS: None

#### PUBLICATIONS AND REPORTS (YEARS 1-2)

- 1) E. Shyamsunder, S.M. Gruner, M.W. Tate, D.C. Turner, P.T.C. So and C.P.S. Tilcock (1988). Observation of inverted cubic phase in hydrated dioleoylphosphatidylethanolamine membranes, *Biochem.* 27: 2332-2336.
- 2) E. Shyamsunder and S.M. Gruner. An x-ray diffraction study of the effects of pressure on  $H_{II}$  phase lipid. (Manuscript in preparation.)

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